

CONDITION REPORT AND TREATMENT PROPOSAL (Mounted Head)

Gallery

SPECIMEN:	Wapiti (Elk) or Red Deer (<i>Cervus elaphus</i>), adult, male.
CATALOG #:	Coe Hall (temp. no. 4; or Elk 1/Elk in Gallery in photo and digital documentation)
PROVENANCE:	Coe Hall at Planting Fields Arboretum State Historic Park, Oyster Bay, New York. Mounted over the archway in the Gallery (1 st floor, F.12) that leads to the Writing Room(1 st floor, F.15). Possibly collected by William Robertson Coe (b.1869) who built Coe Hall, or his son, William Rogers Coe (b. 1901), ca. 1920-24. Coe Hall was a private home until 1949, when it was deeded to the State of New York. In 1955, the State University of New York obtained use of the site as a horticultural study campus and the house was used for classrooms and administration. Some furnishings, including taxidermy specimens, apparently remained in place during this period, and from 1970-78, when the building was used as a community center. In 1978, restoration of Coe Hall was begun, and it has been operated as an historic site since that time.
STATUS:	<i>C. elaphus</i> occur in Europe, parts of Asia, southern Siberia, Mongolia, Manchuria, Korea, parts of China, northwestern Africa, southern Canada, and parts of the US. Populations in many parts of the world are greatly reduced, and at least 2 US subspecies are extinct. The North American population is now subject to controlled sport hunting.
PREPARATION:	Mounted head and antlers.
DIMENSIONS:	D ~98 cm W ~150 cm H not measured.
DESCRIPTION:	<i>(Specimen examined in situ, in ambient light with auxiliary illumination from a flashlight)</i>
Specimen	Taxidermy mount of head and 13 point antlers with soft tissues of nose and the eyelines modeled in wax. Skin shaped over unidentified form. Mouth closed. Metal earliners, possibly lead. Glass eyes, probably painted on reverse. Mane on neck not very heavy.
Plaque	None.
Labels	Unidentified octagonal tag, metal alloy, stamped, "8778" suspended by iron alloy wire with lead (?) seal, from L antler.
CONDITION:	<i>(Specimen examined in situ, in ambient light with auxiliary illumination from a flashlight)</i>
General Specimen	Good. Dusty overall.
Hair	Dark pigments faded. Long hair at base of throat still flexible. There may have been some hair losses from the mane.
Skin	Generally good.
Eyes	Glass dusty. Small cracks and small skin voids on eyelines.
Ears	Exfoliation of skin and some tears inside L ear, and tear near base of ear.. Earliners corroded. Tears at tips of both ears. Long tear inside R ear.
Nose	Cracked across nostrils. Skin lifting slightly

<i>Mouth</i>	Crack on chin, just below lower lip.
<i>Teeth</i>	Not applicable.
<i>Antlers</i>	Very dusty, otherwise good.
<i>Neck</i>	Crack in skin at throat, just below chin.
Plaque	Not applicable.
Labels	Minor corrosion on metal tag and lead (?) seal. Wire very corroded.
ANALYSES:	Testing for the presence of arsenic (see attached for discussion of test procedure) produced a positive result for this specimen.

PROPOSED TREATMENT:

1. Clean specimen using HEPA filtered vacuum. Vacuum through a screen to protect hair.
2. Clean antlers, glass eyes, and wax and painted areas using 95% ethanol on lightly dampened swabs or polyvinyl alcohol sponges, after testing to determine the solubility of the paints (paints of this era when the specimen was likely to have been mounted should be fairly stable to ethanol).
3. Clean metal tag and wire using 95% ethanol.
4. Reattach lifting skin using B-72 (ethyl methacrylate/methyl acrylate copolymer resin) as 50% solids in reagent grade acetone with a small amount of 95% ethanol added (acetone soluble when dry).
5. Infill small skin voids using either B-72 (as above) bulked with Cabosil (fumed silica) or A4M (4,000 mol. wt.) grade methyl cellulose bulked with L tissue and mulberry tissue (softens in water and/or ethanol/water mixtures when dry).
6. Coat wire on metal tag using B-72 ~10% solids in reagent grade acetone (acetone soluble when dry).
7. Tint infills using Daler Rowney acrylic paints with high lightfastness, thinned in water (ethanol soluble when dry).
8. Check the base of the form for labels or other information and if found, record photographically or digitally.
9. Clean base by vacuuming and then with solvents as testing may show to be appropriate.
10. Prepare a black Volara 2A (irradiation crosslinked, expanded polyethylene foam with carbons black filler) pad to cushion the specimen while on display, making sure the pad does not obscure any label data on the base. Adhere to form using 3M #415 doubled-sided tape (acrylic adhesive).

TREATMENT RATIONALE:

By reattaching the skin with a strong but still moderately extensible adhesive, there may be less chance for additional distortion and tearing of the skin as a response to humidity fluctuations in the future. This will also make it possible for staff to undertake routine maintenance on the specimen without further jeopardizing the skin. Repairs will improve the visual appearance of the mount, but will not make it appear as it did originally (e.g., hair will not generally be recolored to try to approximate original coloration). Careful examination will provide an opportunity to recover

additional information about the history of the specimen. All materials to be used in the proposed treatments are synthetics and could not be mistaken for original specimen or original specimen preparation materials in x-radiography or other analysis, and should not interfere with biochemical analysis of untreated areas of the skin.

PHOTOGRAPHY:

Before treatment

(October 2000) ASA 200 Kodak color prints (see photography records, filed by roll and negative number) and digital images (attached).

Roll 2:07, 13-17

REFERENCES:

Down, J., M. MacDonald, J. Tetreault, and R. Williams. 1992. *Adhesive Testing at the Canadian Conservation Institute - An Evaluation of Selected Poly(vinyl acetate) and Acrylic Adhesives*. Environment and Deterioration Report No. 1603. Canadian Conservation Institute, Ottawa.

Nowak, R. and J. Paradiso. 1983. *Walker's Mammals of the World*. 4th ed. The Johns Hopkins University Press. Baltimore and London.

Ruehrwein, R. 1997. Coe Hall at Planting Fields Arboretum State Historic Park. The Creative Company, Lawrenceburg, Indiana.

ESTIMATE:

~ 8 hours, including written, digital, and photographic documentation of the work (2 hours). The estimate assumes that the specimen will be removed from the wall, but will be treated on site, and that no more than 1 year has elapsed between preparation of this report and the actual treatment.

ADDITIONAL CONDITION/TREATMENT NOTES:

TREATMENT APPROVED: _____

(legal custodian or authorized agent)

DATE: _____

TREATMENT REPORT (Mounted Head)

SPECIMEN: Wapiti (Elk) or Red Deer (*Cervus elaphus*), adult, male.

CATALOG #: Coe Hall (temp. no. 4; or Elk 1/Elk in Gallery in photo and digital documentation)

PROVENANCE: Coe Hall at Planting Fields Arboretum State Historic Park, Oyster Bay, New York. Mounted over the archway in the Gallery (1st floor, F.12) that leads to the Writing Room (1st floor, F.15). Possibly collected by William Robertson Coe (b.1869) who built Coe Hall, or his son, William Rogers Coe (b. 1901), ca. 1920-24. Coe Hall was a private home until 1949, when it was deeded to the State of New York. In 1955, the State University of New York obtained use of the site as a horticultural study campus and the house was used for classrooms and administration. Some furnishings, including taxidermy specimens, apparently remained in place during this period, and from 1970-78, when the building was used as a community center. In 1978, restoration of Coe Hall was begun, and it has been operated as an historic site since that time.

On 16 September 2002, the specimen was carefully removed from the wall by Marshall Fine Arts of Deer Park, NY, and placed, hanging upright, on a wood support for treatment.

Based on label found on specimen (see below) after removal from wall, taxidermy was the work of James Lippitt. Clark (1883-1969), a well-known taxidermist and artist who worked with Carl Akeley at the American Museum of Natural History and had a private studio in New York. He prepared specimens for Theodore Roosevelt and other prestigious clients.

STATUS: *C. elaphus* occur in Europe, parts of Asia, southern Siberia, Mongolia, Manchuria, Korea, parts of China, northwestern Africa, southern Canada, and parts of the US. Populations in many parts of the world are greatly reduced, and at least 2 US subspecies are extinct. The North American population is now subject to controlled sport hunting.

PREPARATION: Mounted head and antlers.

DIMENSIONS: D ~42" W ~48" H ~62" (approximate)

DESCRIPTION: *Specimen examined in situ, in ambient light with auxiliary illumination from a flashlight, 17 October 2000 (see Condition Report). Additional descriptive information noted during cleaning and treatment under Ott lamps and ambient light, 2002.*

Specimen Taxidermy mount of head and 13 point antlers with some soft tissues of nose and the eyelines modeled in wax. Skin shaped over wood form with some plaster modeling. Mouth closed. Metal earliners, possibly lead. Glass eyes, probably painted on reverse. Iron alloy hanging hardware on base of form.

Plaque None.

Mounting hardware Ferrous metal, possibly painted. Attached to top edge of wooden form.

Labels	Unidentified octagonal tag, metal alloy (possibly copper alloy or plated with a copper alloy), stamped, "8778" suspended by iron alloy wire with lead (?) seal, from L antler. Seal is embossed, "WYO./GAME/TAG on obverse. Plated metal label on neck end of wooden form is embossed, "MOUNTED BY/JAMES L. CLARK/NEW YORK" and is attached to the form with 2 small brads.
CONDITION:	<i>Specimen examined in situ, in ambient light with auxiliary illumination from a flashlight, 17 October 2000 (see Condition Report). Additional condition information noted during cleaning and treatment under Ott lamps and ambient light, 2002.</i>
General	Fair. Very dusty overall, especially on dorsal surfaces.
Specimen	
<i>Hair</i>	Dark pigments faded. Long hair at base of throat still flexible. There may have been some hair losses from the mane and general loss of some guard hair. Hair has gray/black sooty deposits on surface.
<i>Skin</i>	Generally good.
<i>Eyes</i>	Glass dusty. Small cracks and small skin voids in eyelines.
<i>Ears</i>	Exfoliation of skin and some tears inside L ear, and tear near interior base of ear. Earliners corroded. Tears at tips of both ears. Long tears in skin inside and on back edge of R ear. Voids in hair at tips of both ears. Long tear along back edge of L ear.
<i>Nose</i>	Cracked across nostrils. Skin lifting slightly. Cracks below nostrils.
<i>Mouth</i>	Skin split on chin, just below lower lip.
<i>Teeth</i>	Not applicable.
<i>Antlers</i>	Very dusty and some gray/black deposits, especially on dorsal surfaces. Small holes at back of base of each antler, possibly nail holes or perhaps old insect damage. Otherwise good.
<i>Neck</i>	Skin split at throat, just below chin.
Plaque	Not applicable.
Mounting hardware	Possibly corroded. Difficult to examine given the position of the mount on the wood support for treatment.
Labels	Minor corrosion on metal tag and lead (?) seal on L antler. Wire oxidized. Metal label on base of neck in good condition.
ANALYSES:	Testing for the presence of arsenic (see separate report for discussion of test procedure) produced a positive result for this specimen.
PROPOSED TREATMENT:	<ol style="list-style-type: none"> 1. Clean specimen using HEPA filtered vacuum. Vacuum through a screen to protect hair. 2. Clean antlers, glass eyes, and painted areas using 95% ethanol on lightly dampened swabs or polyvinyl alcohol sponges, after testing to determine the solubility of the paints (paints of this era when the specimen was likely to have been mounted should be fairly stable to ethanol). 3. Clean metal tag and wire using 95% ethanol.

4. Reattach torn skin using B-72 (ethyl methacrylate/methyl acrylate copolymer resin) as 50% solids in reagent grade acetone with a small amount of 95% ethanol added (acetone soluble when dry).
5. Infill small skin voids using either B-72 (as above) bulked with Cabosil (fumed silica) or A4M (4,000 mol. wt.) grade methyl cellulose bulked with L tissue and mulberry tissue (softens in water and/or ethanol/water mixtures when dry).
6. Coat wire on metal tag using B-72 ~10% solids in reagent grade acetone (acetone soluble when dry).
7. Tint infills using Daler Rowney acrylic paints with high lightfastness, thinned in water (ethanol soluble when dry).
8. Check the base of the form for labels or other information and if found, record photographically or digitally.
9. Clean base by vacuuming and then with solvents as testing may show to be appropriate.
10. Prepare a black Volara 2A (irradiation crosslinked, expanded polyethylene foam with carbons black filler) pad to cushion the specimen while on display, making sure the pad does not obscure any label data on the base. Adhere to form using 3M #415 doubled-sided tape (acrylic adhesive).

TREATMENT RATIONALE:

By reattaching the skin with a strong but still moderately extensible adhesive, there may be less chance for additional distortion and tearing of the skin as a response to humidity fluctuations in the future. This will also make it possible for staff to undertake routine maintenance on the specimen without further jeopardizing the skin. Repairs will improve the visual appearance of the mount, but will not make it appear as it did originally (e.g., hair will not generally be recolored to try to approximate original coloration). Careful examination will provide an opportunity to recover additional information about the history of the specimen. All materials to be used in the proposed treatments are synthetics and could not be mistaken for original specimen or original specimen preparation materials in x-radiography or other analysis, and should not interfere with biochemical analysis of untreated areas of the skin.

TREATMENT APPROVAL: Treatment approval signed by Ellen Cone Busch on 16 September 2002.

ACTUAL TREATMENT:

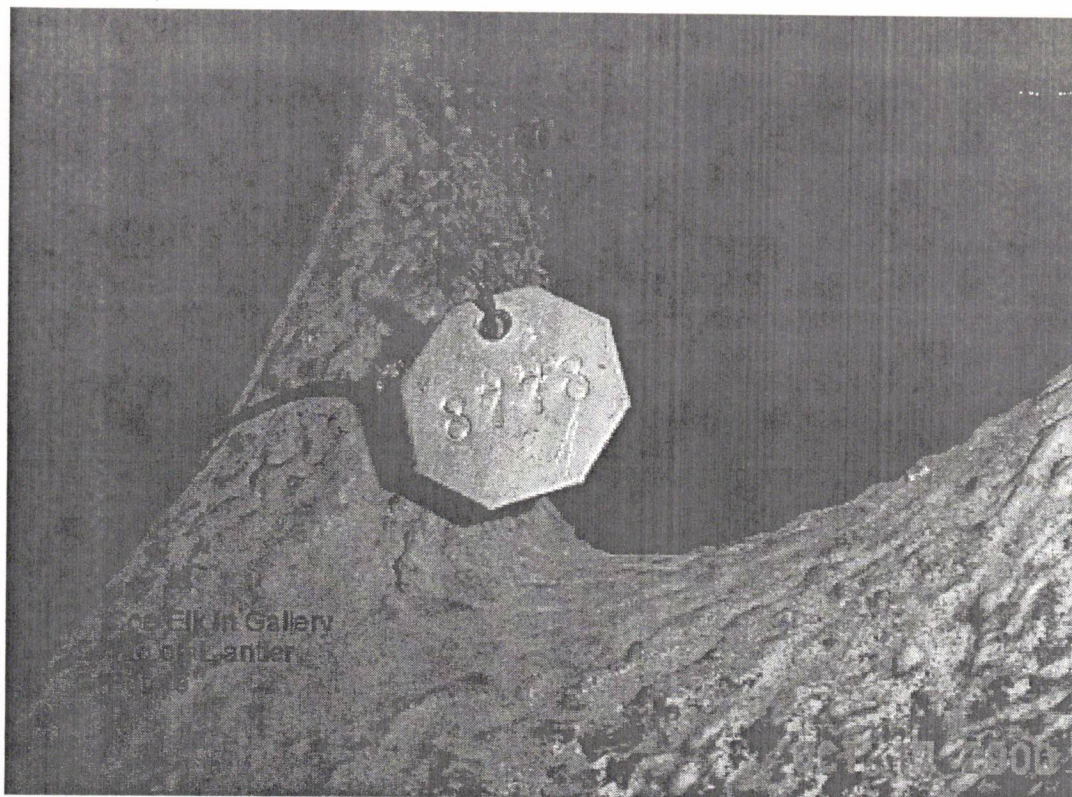
Cleaning

1. Vacuumed with a HEPA filtered vacuum, equipped with small brush as the nozzle.
2. Cleaned hair surface with ethanol/water solution on microfiber cleaning cloths and Kimwipes.
3. Cleaned antlers using soot removal sponges, then with 50% ethanol/water solution on swabs and Kimwipes.
4. Cleaned wire, metal tag, and seal on L antler using 95% ethanol.
5. To degree possible, cleaned metal earliners with 95% ethanol.
6. To degree possible, cleaned ferrous metal hanging hardware using 95% ethanol.

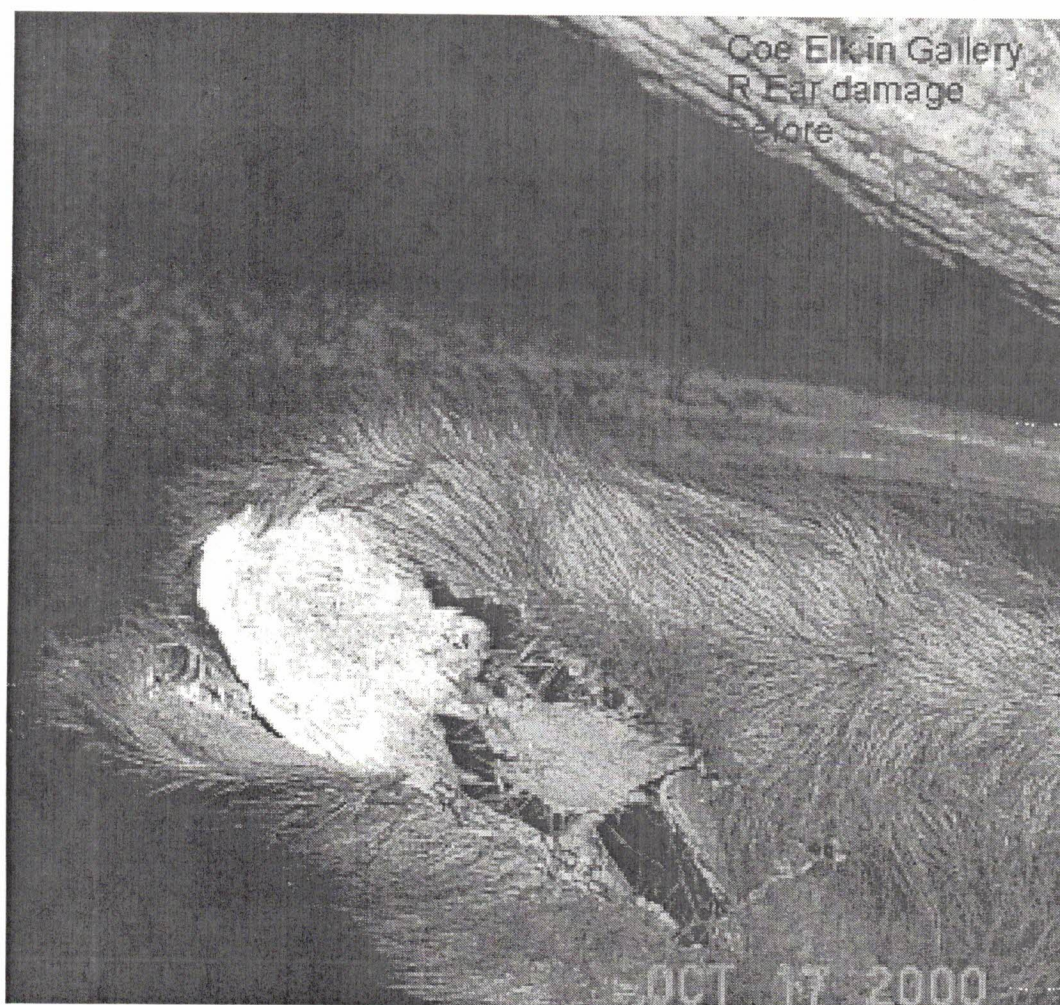
Repairs	<ol style="list-style-type: none"> 1. Coated hanging hardware (to degree possible), and wire, seal and metal tag on L antler using a thin solution of B-72 (ethyl methacrylate/methyl acrylate copolymer resin) solids in reagent grade acetone with a small amount of 95% ethanol added (acetone soluble when dry). Repeated coating. 2. Re-attached skin on mouth, chin, and nose using a 50% solution of B-72 (ethyl methacrylate/methyl acrylate copolymer resin) solids in reagent grade acetone, bulked with fumed silica (Cabosil). 3. To degree possible, coated metal earliners with a thin solution of B-72 in acetone. 4. Made fills in eyelines and nose using a 50% solution of B-72 (ethyl methacrylate/methyl acrylate copolymer resin) solids in reagent grade acetone, bulked with fumed silica (Cabosil). 5. Inside R ear, made a support under the folds of the skin using white Volara 2A foam attached with HMG adhesive (a fairly thick solution of B-72 in acetone), then attached skin to Volara 2A using the same adhesive. Reattached skin to itself at split along back edge of ear using the same adhesive. Made minor infills inside ear using methylcellulose (4,000 molecular weight, A4M) as a gel in deionized water, bulked with alpha cellulose fibers (L tissue fibers). 6. Made infills across nose and inside nostrils using methylcellulose (4,000 molecular weight, A4M) as a gel in deionized water, bulked with alpha cellulose fibers (L tissue fibers). 7. Reattached skin to itself along back edge of L ear and at tip of ear. 8. Made infills at tips of both ears and along back edge of L ear using synthetic hair (National Fiber Technologies no. 13) attached with 50% B-72 in acetone. 9. Tinted all infills using acrylic paints (Liquitex and Daler Rowney) mixed in deionized water (ethanol soluble when dry).
Other	Left black Volara 2A and 3M 415 double-sided tape for use in making pads at base wooden form in each mount when Marshall Fine Arts arrives to replace the trophies on the walls in Coe Hall.
PHOTOGRAPHY:	<p><i>October 2000.</i> ASA 200 Kodak color prints (see photography records, filed by roll and negative number) and images.</p> <p><i>September 2002.</i> ASA 200 Ektachrome slides and 200 ASA Kodak color prints taken with a Nikon autofocus camera and digital images taken with a Sony Cybershot camera.</p>
Before treatment	<p>Roll 02: 07, 13-17 04: 30-34, 36</p> <p>Images: Coe 1 2002: 1607 (label on base of neck)</p>
During treatment	Images: Coe 1 2002: 1609 (seal and label after cleaning and before coating); 1612 (unpainted fills on L ear); 1614 (unpainted fill on R ear); 1615 (unpainted fills on nose)
After treatment	<p>Roll 05: 02-05</p> <p>Images: Coe 1 2002: 1619 (L eye); 1620 (L ear); 1622 (R eye); 1624 (nose); 1625 (L side); 1626 (R side); 1627 (tags on L antler)</p>

REFERENCES:

- Clark, J.L. 1966. *Good Hunting: Fifty Years of Collecting and Preparing Habitat Groups for the American Museum*. University of Oklahoma Press, Norman.
- Down, J., M. MacDonald, J. Tetreault, and R. Williams. 1992. *Adhesive Testing at the Canadian Conservation Institute - An Evaluation of Selected Poly(vinyl acetate) and Acrylic Adhesives*. Environment and Deterioration Report No. 1603. Canadian Conservation Institute, Ottawa.
- Nowak, R. and J. Paradiso. 1983. *Walker's Mammals of the World*. 4th ed. The Johns Hopkins University Press. Baltimore and London.
- Ruehrwein, R. 1997. *Coe Hall at Planting Fields Arboretum State Historic Park*. The Creative Company, Lawrenceburg, Indiana.



Coe Elk in Gallery
R Ear damage
Before



Coe Elk in Gallery
R Ear damage
Before



Label on base of wooden form, Elk (no. 04), Coe Hall