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A "Transparent Chocolate Agar" for the Primary Isolation of
the Neisseria and Hemoglobinophiles.

Reference: Steinberg, P. and Mollov, M. J. Lab. Clin. Med. 27:656-659, Feb. '42.

Directions: Stock agar base. Suspend 1 g. of soluble starch in 1,000 c.c. of cold, distilled water. Mix well. Add 45 g. of proteose No. 3 agar (Difco) and mix. Boil for 1 to 2 min. to dissolve the agar and starch. Distribute to test-tubes in 20 to 40 c.c. amounts, or as desired. Sterilize in autoclave for 20 min. at 15 lbs. pressure.

Preparation of final medium. For every 20 c.c. of melted stock agar base, which has been cooled to about 50°C., add 1 c.c. of sterile defibrinated or citrated blood, (human may be used). Mix well. Heat mixture in water bath to 80°C. to 85°C., and allow to remain at this temperature until blood begins to coagulate. Centrifuge at 2,000 r.p.m. for 1 to 2 min.. Pour supernatant fluid into a Petri dish or prepare slants.